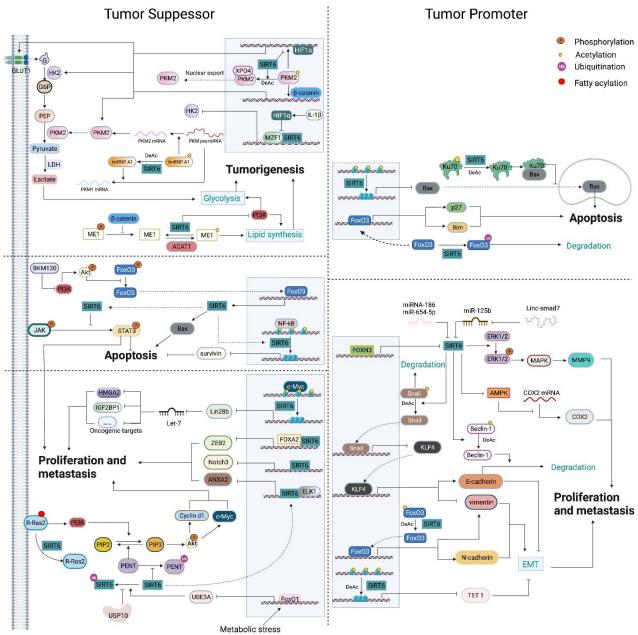
SIRT6 in Aging, Metabolism, Inflammation and Cardiovascular Diseases

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Supplementary Figure 1. SIRT6 regulates the onset and development of cancers SIRT6 inhibits expression of downstream target genes of HIF1 α , such as PDK1, LDH, HK2 and PKM2, thereby suppressing glycolysis process. SIRT6 deacetylates PKM2 at K433 to ensure its interaction with XPO4, thereby promoting PKM2 nuclear export and abolishing its functions as transcriptional coactivator of β -catenin. SIRT6 also deacetylates hnRNP A1 at four sites (K3, K52, K87, and K350) to block alternative splicing of PKM mRNA to PKM2, thereby inhibiting PKM2 expression and PKM2- β -catenin signaling pathway. The SIRT6-MZF1 complex on HK2 promoter negatively regulates HK2, while IL-1 β -induced HIF-1 α abolishes the interaction between SIRT6 and MZF1. SIRT6 deacetylates ME1 at K337 to promote its activity and subsequent lipid synthesis, eventually inhibiting CRC tumorigenesis.

BKM120 inhibits PI3K/Akt signaling to decrease the phosphorylation of FoxO3a, promoting its accumulation on SIRT6 promoter and triggering SIRT6 transcription. On the promoter of survivn, SIRT6 deacetylates H3K9 and reduces NF-κB accumulation to

inhibit survivn expression, thereby promoting cancer cells apoptosis. SIRT6 also activates Bax and suppresses JAK/STAT3 signaling pathway to enhance cellular apoptosis. In some cancer cells, however, SIRT6 deacetylates H3K9 within Bax promoter to inhibit Bax expression. SIRT6 also deacetylates Ku70 at K542 to facilitate association between Ku70 and Bax, which blocks Bax translocation to mitochondria and thereby inhibiting cancer cells apoptosis. SIRT6 ubiquitinates FoxO3 to reduce FoxO3 translocation into the nucleus, thereby reducing expression of FoxO3 target genes p27 and Bim and inhibiting cellular apoptosis.

UBE3A ubiquitylates SIRT6 at K160 to promote SIRT6 degradation. Under metabolic stress, FOXO1 transcriptionally represses UBE3A to stabilize SIRT6. Through deacetylating H3K9, SIRT6 acts as a corepressor of ELK1 to suppress ANXA2 expression, thereby reducing the proliferative capacity of cancer cells. SIRT6 inhibits Notch3 signaling pathway to inhibit cellular proliferation and invasion. Coordination of FOXA2 and SIRT6 on the promoter of ZEB2 inhibits its transcription, which resulting in decreased cancer cell progression. SIRT6 also prevents PENT ubiquitylation and degradation, decreasing phosphorylation of PIP3 to form PIP2, inhibiting PI3K/Akt pathway and downstream genes expression such as c-myc and cyclin d1, and eventually repressing cellular proliferation. USP10 stabilizes SIRT6 to antagonize the transcriptional activity of c-myc. SIRT6 also inhibits plasma membrane localization of R-Ras2 through reducing its lysine fatty acylation at four sites (K192, K194, K196, and K197), thereby inactivating PI3K/Akt signaling pathway. SIRT6 represses c-myc-driven transcription of lin28b through deacetylating H3K9 and H3K56, thereby abolishing the suppression of lin28b on Let-7. Let-7 promotes the degradation of oncogenic proteins, such as insulin growth factor 2 binding proteins (IGF2BPs) and high mobility group AT-hook 2 (HMGA2), inhibiting cellular growth.

SIRT6 could upregulate phosphorylation of ERK1/2 to activate MAPK-MMP9 pathway, enhancing cancer invasiveness. FOXN3 downregulates SIRT6 transcription. MiR-186 and miR-654-5p both impair SIRT6 expression to reduce proliferation capacity of cancer cells. On the contrary, Linc-smad7 sponges miR-125b to upregulate SIRT6 expression, promoting cellular proliferation and invasion. SIRT6 downregulates AMPK activity to stabilize mRNA of COX-2, thereby enhancing COX-2 translation and cellular proliferation. SIRT6 deacetylates Snail to prevent its degradation, and then Klf4 expression is suppressed. Klf4 maintains E-cadherin expression and reduces vimentin expression, thereby inhibiting EMT. SIRT6-mediated deacetylation of Beclin-1 promotes autophagic degradation of E-cadherin. SIRT6 deacetylates FoxO3a to promote N-cadherin and Vimentin expression, thereby enhancing EMT. Conversely, through deacetylating H3K9, SIRT6 suppresses TET 1 transcription to accelerate EMT.

1. SIRT6 regulates the onset and development of cancers

In many human cancers, downregulation of SIRT6 expression was reported, which in a large part was associated with increased tumor progression and poor clinical outcome. However, other studies also found that SIRT6 could exert oncogenic functions in a subset of human cancers¹. Therefore, SIRT6 was considered as a double-edged sword that possesses dual roles of tumor suppressor and promoter (Supplementary Fig. 1).

1.1 Tumorigenesis

Aerobic glycolysis known as the Warburg effect is an important source for energy and macromolecular synthesis in the development and progression of cancer cells. As mentioned above, SIRT6 acts as a corepressor of HIF1α to inhibit glycolysis². In SIRT6-deficient cells, expression levels of glycolytic genes such as PDK1, LDH, hexokinase 2(HK2) and PKM2, were enhanced, and increased aerobic glycolysis promotes oncogenic transformation in an oncogene activation-independent manner³. On the upstream, FoxO3a and RUNX2 could upregulate and downregulate SIRT6 transcription, respectively, to regulate glycolysis and subsequent cell differentiation, viability, and clinical outcome^{4,5}. As a key rate-limiting enzyme of aerobic glycolysis, the overexpression of PKM2 contributes to tumorigenesis⁶. SIRT6 was shown to deacetylate heterogeneous nuclear ribonucleoproteins A1(hnRNP A1) to block alternative splicing of PKM mRNA to PKM2, reducing PKM2 expression⁷. In addition, deacetylation of PKM2 by SIRT6 led to the nuclear export of PKM2, thereby abrogating its non-glycolytic functions (i.e., nuclear protein kinase and transcriptional coactivator functions)8. SIRT6 was also shown to be a suppressor of HK2, regulating inflammation and glycolysis in the tumor microenvironment⁹. Inhibition of PI3K signaling by SIRT6 at the transcriptional level suppresses glycolysis and lipid metabolism in cancer stem cells, antagonizing tumor sphere formation 10. In addition, SIRT6 antagonizes acetyl-CoA acetyltransferase (ACAT1) function to inhibit malic enzyme 1 (ME1) activity, which reduces NADPH for fatty acid biosynthesis, eventually suppressing colorectal tumorigenesis¹¹. However, in papillary thyroid cancer cells, SIRT6 was shown to upregulate glycolytic genes expression and Warburg effect via upregulation of ROS¹².

1.2 Proliferation and metastasis of cancer cells

Uncontrolled cellular proliferation is one of the hallmarks of cancer progression. Upon metabolic stress, FoxO1 transcriptionally represses the expression of E3 ubiquitin ligase UBE3A to inhibit degradation of SIRT6 by UBE3A, which, in turn, results in Annexin A2 (ANXA2) repression, eventually reducing the proliferative capacity and invasiveness of HCC¹³. In colon cancer, USP10 stabilizes SIRT6 to antagonize the transcriptional activity of the c-myc, and then induces cell cycle arrest and tumor growth inhibition¹⁴. In addition, SIRT6 increases expression and stability of PENT to disrupt PI3K/AKT signaling, suppressing proliferation and invasion of colon cancer cells¹⁵. SIRT6 also decreases the lysine fatty acylation of R-Ras2 to reduce its membrane localization and subsequent activation of PI3K/AKT signaling, suppressing cellular proliferation¹⁶. In addition, SIRT6 downregulation is associated with poor prognosis of pancreatic ductal adenocarcinoma (PDAC)¹⁷. Loss of SIRT6 results in hyperacetylation of histone and cmyc recruitment within the Lin28b promoter to upregulate expression of Lin28b and downstream let-7 target genes, promoting proliferation and progression of PDAC¹⁷. Downregulation of SIRT6 by miR-34c-5p enhances cell proliferation through activating JAK2/STAT3 signaling pathway¹⁸. In addition, SIRT6 represses proliferation and invasion through inhibiting Notch3 signaling pathway^{19,20}. SIRT6 also positively regulates the levels of the phosphorylated extracellular signal-regulated kinases 1 and 2 (pERK1/2) to activate matrix metalloproteinase-9 (MMP-9) signaling pathway, promoting cell growth and metastasis in bone cancer²¹. However, forkhead box N3 (FOXN3) transcriptionally inhibits the SIRT6 expression, thereby repressing MMP9 to downregulate proliferation in osteosarcoma²². In human squamous cell carcinoma, UVB upregulates SIRT6 expression via activating the AKT pathway, and then SIRT6 promotes the expression of COX-2, increasing cell survival and proliferation²³. Downregulated SIRT6 by miRNA-186 and miR-654-5p contribute to impaired proliferation capacity of cancer cells^{24,25}.

1.3 Epithelial-mesenchymal transition of cancer cells

Epithelial–mesenchymal transition (EMT) confers malignant properties, such as invasiveness and metastasis, to carcinoma cells²⁶. SIRT6-mediated invasiveness is significantly related to the expression of EMT associated molecules, such as E-cadherin, N-cadherin, vimentin, Snail and activated β-catenin^{27,28}. Mechanistically, SIRT6 not only deacetylates Beclin-1 to enhance the autophagic degradation of E-cadherin, but also deacetylates FoxO3 to promote N-cadherin and vimentin expression, eventually promoting EMT of HCC²⁹. In addition, long intergenic noncoding RNA smad7 (Linc-smad7) upregulates SIRT6 through sponging miR-125b, promoting EMT of HCC³⁰. SIRT6 also promotes the deacetylation and stabilization of Snail to repress the expression of *Klf4*³¹. Subsequently, reduced KLF4 results in decreased E-cadherin expression and increased vimentin expression, promoting EMT and aggressiveness of non-small cell lung cancer³¹. In colon carcinoma cells, SIRT6 suppresses tet methylcytosine dioxygenase 1 (TET 1) transcription through reducing H3K9 deacetylation to accelerate EMT³². However, in PDAC cells, transcriptionally upregulated expression of SIRT6 by KLF10 was shown to ameliorate glycolysis, EMT and metastasis³³. In addition, coordination of SIRT6 and FOXA2 inhibits the expression of zinc finger E-box binding homeobox 2 (ZEB2) to suppress proliferation and invasion of HCC³⁴.

1.4 Apoptosis of cancer cells

Cellular apoptosis is an important part of tumor development, therefore, therapies triggering apoptosis have been used to eliminate malignant cells³⁵. Early animal studies found that SIRT6 could inhibit the liver cancer development at the initiation stage through reducing the antiapoptotic activity of survivn via reducing both H3K9Ac and NF-κB levels at the survivin promoter³⁶. Furthermore, the mono-ADP ribosyltransferase activity of SIRT6 also potentiates apoptosis in cancer cells line through ATM-mediated p53 and p73 signalling cascades, but not in normal cells³⁷. Interestingly, unlike SIRT6 in proliferation, SIRT6 was shown to block the ERK signaling pathway to induce cancer cell apoptosis³⁸. In glioma cells, SIRT6 induces apoptosis through activating the JAK2/STAT3 signaling pathway³⁹. In colorectal cancer cells, PI3K inhibitor BKM120 was found to positively regulate SIRT6 expression by reducing phosphorylation level of FoxO3a, which in turn promotes apoptosis through activating Bax and mitochondrial pathway⁴⁰. However, SIRT6 could also play oncogenic roles through attenuating Bax signaling to potentiate apoptosis evasion in HCC^{41,42}. Mechanistically, on one hand, SIRT6 could inhibit Bax expression through deacetylating H3K9; on the other hand, SIRT6 deacetylates Ku70 to promote Bax-Ku70 interaction, thereby blocking Bax mitochondrial translocation^{41,42}. In addition, SIRT6 prevents FoxO3 translocation into the nucleus, which reduces expression of its target genes p27 and Bim, thereby preventing doxorubicin-induced cell death⁴³.

Collectively, SIRT6 fulfills a controversial role in regulating complex network of biological pathways in different cancer cells, which is partly due to the high heterogeneity of tumor cells. In general, SIRT6 protects against tumorigenesis, while roles of SIRT6 in cancer are difficult to identified after tumor formation. Though this pleiotropism of SIRT6 adds a layer of difficulty in understanding the cellular mechanisms by which SIRT6 impacts biological processes of cancer cells, opportunities appear to be encouraging for attenuating cancer development through targeting SIRT6. Several studies have shown that the sensitivity of tumor cells to chemotherapy or immunotherapy can be improved through inhibiting SIRT6⁴⁴⁻⁴⁷. Therefore, targeting SIRT6 seems a novel and promising strategy to improve cancer treatment and patient outcome.

2. Neurodegenerative diseases

Alzheimer's disease (AD) and Parkinson's disease (PD) are common ageing-related diseases. AD is involved in neurodegeneration, and characterized by the deposition of Amyloid β (A β) and the accumulation of hyperphosphorylated Tau at the cellular level⁴⁸. SIRT6 reduction was significantly observed in both AD mouse model and patients, implying that the aberration of SIRT6 is closely associated with pathologies of AD^{49,50}. In detail, SIRT6 could prevent A β 42-induced DNA damage, attenuating development of AD⁵⁰. In addition, SIRT6 deficiency results in glycogen synthase kinase 3 (GSK3) activation to increase stability and phosphorylation of Tau protein, a critical mark in neurodegenerative diseases⁴⁹. SIRT6 also deacetylates Tau at K174 to decrease its stability and nuclear accumulation, attenuating DNA damage and nucleolar dysfunction⁵¹. A recent study shows that bolstering neuronal NAD⁺ levels could increase SIRT6 activity, which improves synaptic dysfunction, and neuronal degeneration through attenuating DNA damage, phosphorylation of Tau (pTau) accumulation and neuroinflammation⁵². However, SIRT6 level was reported to be greater in PD patient brains, and in which, SIRT6 plays pathogenic roles through upregulating the proapoptotic TNF- α pathway and downregulating the pro-survival AKT signalling⁵³. In addition, a recent study shows that mRNA level of SIRT6 was up-regulated in the peripheral blood of PD patients⁵⁴, suggesting that SIRT6 may play a pathogenic role in PD development.

In sum, current studies on SIRT6 in neurodegenerative diseases are limited, and SIRT6 seems to play contradictory roles in AD and PD. This discrepancy may be contributed to cell specificity. But in any case, SIRT6 is undoubtedly involved in neurodegenerative diseases, therefore, more research for understanding its specific mechanisms is needed.

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